

Dorsal Raphe Neurons Signal Reward through 5-HT and Glutamate

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<http://dx.doi.org/10.1016/j.neuron.2014.02.010>

SUMMARY

The dorsal raphe nucleus (DRN) in the midbrain is a key center for serotonin (5-hydroxytryptamine; 5-HT)-expressing neurons. Serotonergic neurons in the DRN have been theorized to encode punishment by opposing the reward signaling of dopamine neurons. Here, we show that DRN neurons encode reward, but not punishment, through 5-HT and glutamate. Optogenetic stimulation of DRN Pet-1 neurons reinforces mice to explore the stimulation-coupled spatial region, shifts sucrose preference, drives optical self-stimulation, and directs sensory discrimination learning. DRN Pet-1 neurons increase their firing activity during reward tasks, and this activation can be used to rapidly change neuronal activity patterns in the cortex. Although DRN Pet-1 neurons are often associated with 5-HT, they also release glutamate, and both neurotransmitters contribute to reward signaling. These experiments demonstrate the ability of DRN neurons to organize reward behaviors and might provide insights into the underlying mechanisms of learning facilitation and anhedonia treatment.

INTRODUCTION

Reward processing is a fundamental function of the brain. Animal and human behaviors are reinforced by reward, and the inability to experience rewarding stimuli is a key feature of depression and schizophrenia in humans (Der-Avakian and Markou, 2012). Studies in the last six decades have identified the brain reward system as an interconnected set of brain structures that are important for reward processing. Within this system, dopamine neurons in the midbrain ventral tegmental area (VTA)

are believed to play pivotal roles (Wise and Rompre, 1989). The dorsal raphe nucleus (DRN) projects extensively to several reward-related brain areas, and this structure is the major source of serotonin (5-HT) in the forebrain (Jacobs and Azmitia, 1992; Vertes, 1991). As the 5-HT system is an important target for the treatment of depression and several other major psychiatric disorders (Mann, 1999), studying the functions of DRN neurons in reward signaling might provide insight into the circuit mechanisms of reward processing and may suggest avenues to treating mental disorders.

While there has been much work on reward circuits in the brain, the roles of DRN neurons in reward signaling have been largely overlooked, likely reflecting a wide range of literature reporting conflicting views on the precise relationship between the activity of DRN neurons and animal reward behavior (Kranz et al., 2010). Although DRN neurons are heterogeneous in their neurotransmitter phenotypes, most studies have been focused on the behavioral effects of changing 5-HT levels, and many studies have proposed 5-HT as an opponent to dopamine's rewarding activities. Decreasing brain 5-HT levels promotes impulsive behavior that should normally be suppressed in response to situations involving stress or punishment (Clarke et al., 2004; Crockett et al., 2009; Tye et al., 1977). Pharmacological studies suggest that 5-HT opposes the action of dopamine in reward-associated tasks and inhibits the reinforcement effects of intracranial electric self-stimulation (Abler et al., 2012; Amit et al., 1991; Di Matteo et al., 2001; Fletcher et al., 1999). In addition, some DRN neurons are activated by aversive cues or negative reward (Li et al., 2013; Ranade and Mainen, 2009; Schweimer and Ungless, 2010). Thus, these previous experiments have supported the notion that DRN neurons may encode punishment and mediate behavioral suppression through the release of 5-HT to counter the dopamine system (Cools et al., 2008; Daw et al., 2002; Dayan and Huys, 2009). However, depleting 5-HT impairs reward processing in humans and animals (Miyazaki et al., 2012; Seymour et al., 2012). Several recent electrophysiological studies report that the activity of subsets of DRN neurons is affected by reward size and delay (Bromberg-Martin et al.,

2010; Inaba et al., 2013; Miyazaki et al., 2011; Nakamura et al., 2008; Ranade and Mainen, 2009), suggesting that the DRN might modulate certain aspects of reward processing as well.

Precise activation of DRN neurons can directly test whether the activity of these neurons signifies reward or punishment. In this study, we selectively expressed the light-sensitive cation channel ChannelRhodopsin2 (ChR2) in DRN neurons (Boyden et al., 2005), using the ePet1-Cre mouse line that has been extensively used to drive gene expression in 5-HT neurons (Scott et al., 2005). Unlike slow and diffusive pharmacological manipulations, optogenetic stimulation enables us to precisely activate neurons in the DRN and overcomes the drawback of stimulating the fibers of passage with electrical stimulation. After finding that stimulation of DRN Pet-1 neurons reinforces behavior and guides animal learning, we performed recordings from behaving mice and observed the selective activation of these neurons during reward-associated tasks. Moreover, we combined whole-cell recordings with behavioral assays using mutant mice to show that DRN Pet-1 neurons require glutamate and 5-HT to mediate reward signaling. These results support the concept of DRN as a reward center in the brain and provide important implications for theories of reward and 5-HT functions.

RESULTS

Optogenetic Activation of DRN Pet-1 Neurons Produces Strong Reward

We used an optogenetic approach to stimulate DRN neurons. The gene encoding ChR2-mCherry or mCherry was selectively targeted into DRN neurons of ePet1-Cre mice using a Cre-dependent AAV viral vector carrying a “double floxed” inverted open reading frame (AAV-DIO) (Figure 1A; Figure S1A available online) (Zhang et al., 2010). The Pet-1 gene encodes a transcription factor selectively expressed in brain 5-HT neurons (Hendricks et al., 2003), and the transgenic ePet1-Cre mouse line is commonly used to label brain 5-HT neurons (Scott et al., 2005). Two weeks after virus infusion into the center of the DRN, ChR2-mCherry was robustly expressed throughout the DRN, and its distribution pattern resembled that of 5-HT (Figures 1B and S1B). Because of the membrane expression of ChR2-mCherry, we injected AAV-DIO-mCherry to label Pet-1 neurons with mCherry and examined labeling efficiency and selectivity using the immunostaining of tryptophan hydroxylase 2 (Tph2), a marker of central 5-HT neurons (Zhang et al., 2004). Overall, $96.9\% \pm 0.4\%$ of Tph2⁺ neurons in the DRN were labeled with mCherry, and $92.4\% \pm 0.7\%$ of all mCherry⁺ neurons exhibited clear Tph2 immunoreactivity ($n = 4,669$ mCherry⁺ neurons from 12 brain sections of three mice) (Figures S1C–S1E). DRN Pet-1 neurons did not express tyrosine hydroxylase (TH), a marker of dopamine neurons (Figures S1F and S1G). Whole-cell recordings from brain slices demonstrated that ChR2-mCherry⁺ neurons in the DRN were reliably activated by blue light pulses to faithfully fire action potentials at a frequency of up to 20 Hz (Figures 1C and S1H–S1M).

We investigated the behavioral effects of stimulating DRN Pet-1 neurons using a method named *intraCranial light* administration in a specific subarea (iClass). In the iClass training sessions, mouse exploration within a marked rectangular subarea of an

open field triggered the delivery of blue light pulses to the DRN through an implanted optical fiber (Figure S1B). Normally, mice avoid the center area and prefer contact with the walls and corners. Based on the efficiency of driving gene expression in 5-HT neurons and the punishment theory of the 5-HT system, we originally predicted that ePet1-DRN^{ChR2} mice would avoid a designated corner area after coupling to light stimulation (473 nm, 15 ms pulses at 20 Hz). Surprisingly, these mice exhibited a strong preference of the stimulation-coupled corner (Figures S2A and S2B).

To assay the potential reinforcement effect more stringently, we examined whether mice could be trained to increase exploration in the center subarea of an open field by overcoming the instinctive avoidance of open space (Figures 1D and S2C). All ePet1-DRN^{ChR2} mice dramatically increased center exploration after the start of iClass training using 5 or 20 Hz light pulses, whereas no such effect was observed in control mice lacking functional ChR2 expression in the DRN (Figures 1E, 1F, and S2D–S2F; Movie S1). Minutes after the onset of the first training session, the center entry frequency and exploration time of ePet1-DRN^{ChR2} were increased 8-fold (Figures 1G–1J). Only 6.6 s of optical stimulation over four entries were required to induce a significant change in the exploratory behavior of ePet1-DRN^{ChR2} mice (Figures S2G and S2H). During the following two days, the exploration frequency and duration in the designated center area were further increased to nearly 12-fold that of the baseline (Figures 1G–1J; Movie S2). Stimulation at 5 Hz produced a significant increase in center exploration, although the center entry numbers were fewer than those induced through 20 Hz stimulation (Figures 1G–1J and S2F). Following one or two 15 min sessions without light stimulation, the ePet1-DRN^{ChR2} mice reduced center exploration and locomotor activity to pretraining levels (Figures 1G–1J, S2I, and S2J), indicating that the behavioral effects of iClass training are extinguishable.

Thus, the results of the iClass experiments suggest that activating DRN Pet-1 neurons positively reinforces behaviors and signals reward. We adapted the two-bottle preference test to compare the reward value conferred by the optogenetic activation of DRN Pet-1 neurons relative to the innate valence of ingested sucrose solutions (Domingos et al., 2011). In this test, mice had a choice of licking two contact lickometers to access liquid from the two bottles, which distributed sucrose and water, separately. Mice without light stimulation preferred sucrose solution, and the reward value of sucrose reached a plateau at a concentration of 5% (Figure 2A). For ePet1-DRN^{ChR2} mice, we coupled DRN stimulation with licking for water, but not for sucrose solution (20 Hz 1 s or 5 Hz 2 s) (Figure 2B). Light coupling at either 5 or 20 Hz allowed water to compete favorably against 0%–2% of sucrose and produced preference scores comparable to those observed with 5% sucrose (Figures 2C and 2D). This result indicates that the optogenetic stimulation of DRN Pet-1 neurons produces a reward value of over 5% sucrose solution.

We used the traditional method of self-stimulation to determine whether the optogenetic activation of DRN Pet-1 neurons could support self-stimulation with an instrumental action (Olds and Milner, 1954). Several recent studies have found that

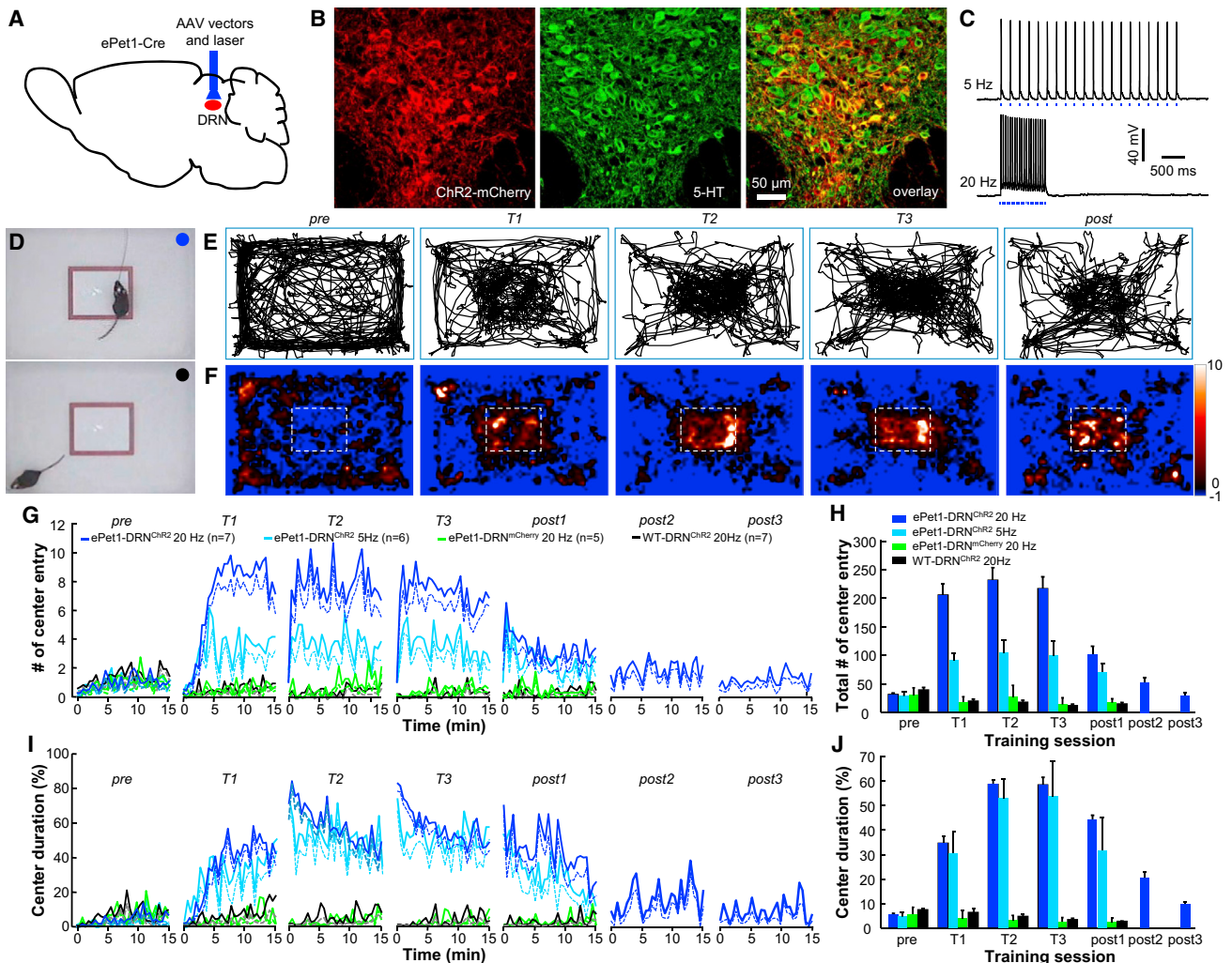


Figure 1. Optogenetic Activation of DRN Pet-1 Neurons Reinforces Area-Specific Exploratory Behavior

(A–C) ChR2 was selectively expressed in DRN Pet-1 neurons by infusing AAV-DIO-ChR2-mCherry viral vectors into the DRN of ePet1-Cre mice (A), which drive ChR2-mCherry expression (red in [B]) in 5-HT neurons (green). Recordings from brain slices demonstrate precise neuronal activation with brief blue light pulses at 5 and 20 Hz (C).

(D) The method of iClass training. The body positions of an ePet1-DRN^{ChR2} mouse were video tracked, and light pulses were delivered to the DRN through an optical fiber when the mouse entered the marked center subarea of an open field (blue circle, upper image). Light was not applied when the mouse was out of the center area (black circle, lower image).

(E and F) The locomotor tracks (E) and heatmaps (F) illustrating the spatial exploration of a mouse before (*pre*), during (*T1–T3*), and after (*post*) iClass training sessions. The color scale at the right indicates the spatial exploration in a specific area normalized by the average time if the mouse had lacked any spatial preference.

(G and H) Plots of the instantaneous rates (G) and the total number (H) of center entries across sessions (30 s per point) for ePet1-DRN^{ChR2} mice, ePet1-DRN^{mCherry} mice, and nontransgenic littermates injected with AAV-DIO-ChR2-mCherry virus (WT-DRN^{ChR2}). The dashed lines indicate mean-SEM. The error bars indicate SEM in this and following figures.

(I and J) The instantaneous ratio (I) and the mean ratio (J) of center duration across sessions. $p < 0.001$; Tukey's multiple comparisons between ePet1-DRN^{ChR2} groups and control groups. See also [Figures S1](#) and [S2](#) and [Movies S1](#) and [S2](#).

operant conditioning could be generated with varying success through the optogenetic stimulation of midbrain dopamine neurons (Adamantidis et al., 2011; Kim et al., 2012; Rossi et al., 2013; Witten et al., 2011). Individual mice were placed in a standard chamber equipped with nose-poke ports (Figure 2E). Nose-poke through the “active,” but not the “inactive,” port caused the passing of brief light pulses to the DRN (20 Hz 3 s or 5 Hz for 2 s), followed by a 5 s timeout period. After 1 hr of conditioning

with either stimulation strength, ePet1-DRN^{ChR2} mice dramatically increased nose poking through the active port (Figure 2F; Movie S3). Stronger stimulation evoked ~700 active pokes and resulted in ~300 stimulations in 1 hr, whereas weaker stimulation generated approximately half of the response intensity (Figures 2G–2I). As a control, there were <10 inactive pokes for ePet1-DRN^{ChR2} mice and a similar small number of active pokes for ePet1-DRN^{mCherry} control mice.

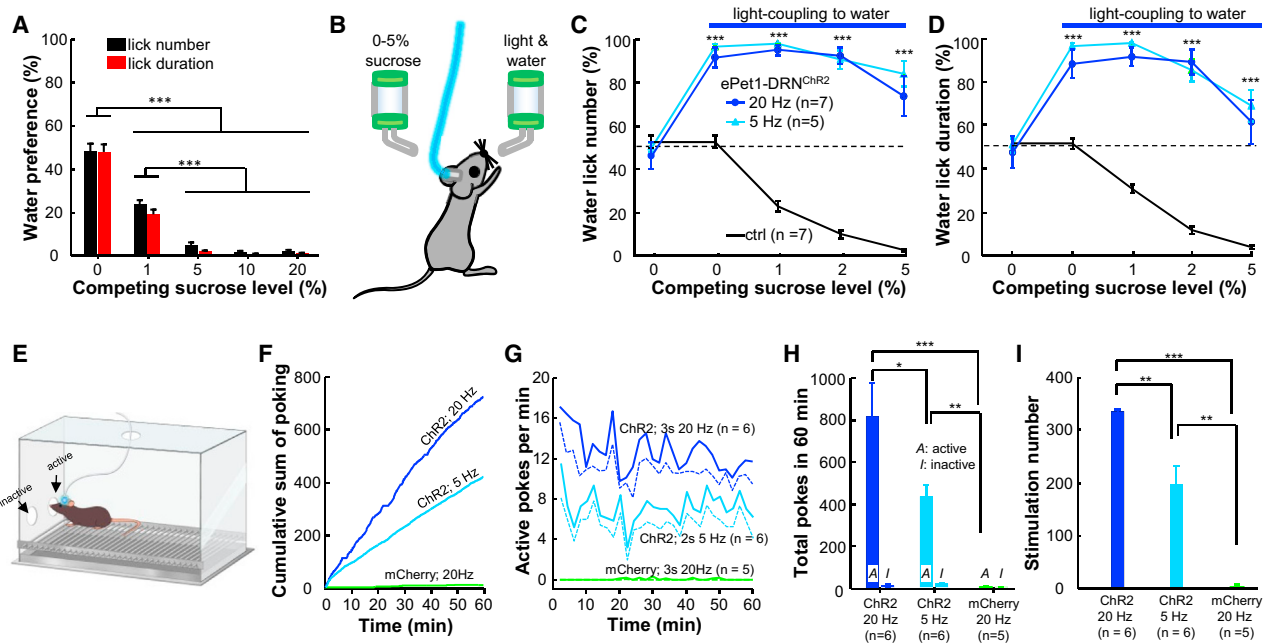


Figure 2. Stimulation of DRN Pet-1 Neurons Shifts Sucrose Preference and Causes Operant Reinforcement for Self-Administration

(A) In two-bottle preference tests, WT mice exhibited a reduced preference for water when the sucrose concentration was increased in the competing bottle. The preference scores were quantified using either lick numbers (black) or lick duration (red). *** $p < 0.01$; one-way ANOVA followed by Tukey's multiple comparisons test; $n = 8$ mice.

(B) The method of testing the effect of DRN neuron activation on shifting sucrose preference.

(C and D) Coupling light stimulation to licking for water increased lick numbers (C) and lick duration (D) for water and shifted animal preference away from sucrose. *** $p < 0.001$; two-way ANOVA with Sidak's multiple comparisons between tests with or without light coupling.

(E–I) DRN stimulation reinforces operant learning.

(E) The method of optical self-stimulation. Mice received DRN light stimulation after nose poking through the “active,” but not the “inactive,” hole of an operant chamber.

(F) Plots of cumulative nose pokes of individual mice. ChR2-expressing mice, but not the mCherry control animals, vigorously poked the “active” hole for self-stimulation.

(G) The rate of active nose pokes across the test sessions of 60 min. ePet1-DRN^{ChR2} mice stably completed ~12 active pokes/min throughout the test sessions with strong light stimulation (3 s; 20 Hz) and approximately seven pokes/min with weaker stimulation (2 s; 5 Hz), whereas the number of active nose pokes was close to zero for the ePet1-DRN^{mCherry} control mice.

(H) Group data showing the total number of active and inactive pokes within a 60 min session.

(I) ePet1-DRN^{ChR2} mice earned more than 300 trains of light stimulation with strong stimulation and ~200 stimulations with weak stimulation, whereas ePet1-DRN^{mCherry} control mice collected only about three stimulations. Due to the 5 s timeout for stimulation delivery, the number of earned stimulations was fewer than that of nose pokes. ** $p < 0.01$; *** $p < 0.0001$; between-group t tests. See also [Movie S3](#).

We further investigated whether the activation of DRN neurons could be used to guide sensory discrimination learning by challenging mice with an olfactory Go/No-go task (Figure 3A). Mice were trained to distinguish between two odors by licking a metal port for reward only after the presentation of an odorant (the odorant is termed CS+, and the action is termed a “hit”). The licking response to another odorant (termed CS– and “false positive,” respectively) resulted in a penalty of timeout. As a control, water-deprived wild-type (WT) mice were trained with 5% sucrose solution as a reward (Figure 3B). These animals took ~500 training trials in 2 days to reach a stable performance of $\geq 90\%$ correct by gradually reducing false positive responses (Figures 3C and 3D). For the ePet1-DRN^{ChR2} mice that were not water deprived, licking after CS+ did not result in fluid release, but rather triggered optical stimulation of the DRN (Figure 3B). These mice performed with high motivation and

accuracy. All test animals completed 500 trials within a single 4 hr training session. The ePet1-DRN^{ChR2} mice reached the initial 90% correct response ratio after only 12 trials and reached a stable performance of $\geq 90\%$ correct after ~30 trials (Figures 3C, S3A, and S3B). Only six CS– trials were needed for the ratio of false positive responses to decrease to 20% (Figures 3D and S3C). After conditioning with the original odorant pair, all ePet1-DRN^{ChR2} mice learned to establish novel associations within ~30 trials (Figures 3E, S3D, and S3E). When the valences of the two conditioning odorants were reversed, these mice learned to adjust their responses in ~100 trials (Figures 3F, S3F, and S3G). As a comparison, the mice trained with sucrose solution completed the switch and reversal learning more slowly (Figure S3H–S3K). Thus, the optogenetic stimulation of DRN Pet-1 neurons efficiently guides the learning of sensory discrimination.

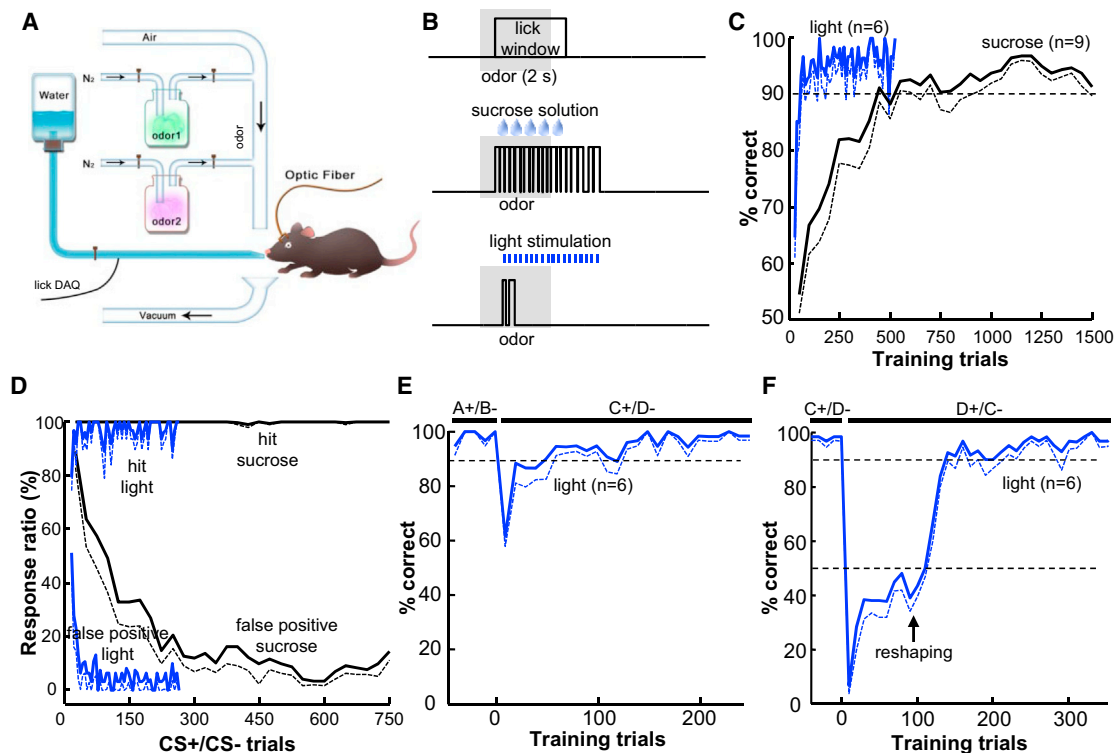


Figure 3. Activation of DRN Pet-1 Neurons Efficiently Guides Sensory Discrimination Learning

(A and B) The method of olfactory Go/No-go tests. Mice learned to lick a touch lickometer for sucrose solution or DRN stimulation in response to one of two odorants (A). The time lines of actions for reward trials are shown in (B). We used light stimulation of the DRN (3 s; 20 Hz) instead of sucrose solution for ePet1-DRN^{ChR2} mice.

(C) The learning curves of odor discrimination for mice trained with the reward of sucrose solution or DRN light stimulation. The dashed curves indicate mean-SEM.

(D) The mean ratio of hit responses to CS+ odor and false positive responses to CS- odor.

(E) Plot of correct ratio of ePet1-DRN^{ChR2} mice in the switch learning phase, during which the original odorant pair (A+/B-) was changed to a novel pair of odorants (C+/D-).

(F) Light stimulation enabled efficient learning of the valence reversal of conditioning odor stimuli (from C+/D- to D+/C-). After odor reversal, the mice abandoned licking in response to both odorants. Sucrose solution was automatically released following the current CS+ odors for 2 or 3 trials, and the licking behavior was "reshaped" for later light stimulation. See also Figure S3.

DRN Pet-1 Neurons Are Activated during Reward-Associated Tasks

We performed multielectrode recordings in behaving mice to examine how DRN Pet-1 neurons respond during reward-associated tasks. The mice were trained to learn Go/No-go olfactory discrimination while head fixed on a floating spherical treadmill (Figure S4A). A delay (1 s) was inserted between the odor cue (1 s) and the time window for sucrose consumption (2 s). After training, the mice reliably showed licking responses during the sucrose delivery time windows after the presentation of the CS+, but not the CS-, odors (Figure S4B). A multichannel optrode was then targeted at the DRN of head-fixed ePet1-DRN^{ChR2} mice engaged in the task (Figures 4A, S4C, and S4D). To address the difficulties of classifying cell types using electrophysiological and pharmacological properties (Kocsis et al., 2006), we identified Pet-1 cells based on the criteria that brief, blue light pulses reliably evoked the firing of action potentials with similar waveforms (Figure 4B).

A majority of 60 identified Pet-1 neurons were significantly activated during reward-associated tasks (Figures 4C and 4D). Unlike midbrain dopamine neurons that are transiently activated after the onset of reward-predictive sensory cues (Cohen et al., 2012; Schultz, 1997), DRN Pet-1 neurons typically fired at approximately five spikes/s before trial onset; increased firing frequency after CS+, gradually reaching an activity peak of ~18 spikes/s during the delay and sucrose consumption periods; and returned to the baseline before the completion of sucrose consumption (Figures 4C, S4E, and S4F). The amplitude and duration of neuronal activation are comparable to the stimulation parameters used for behavioral assays. We calculated the receiver operating characteristic (ROC) value of the neural activity by comparing the spike firing rates after trial onset to those in a control time window before trial onset (Figure S4G) (Nakamura et al., 2008). The response strength was then quantified as the area under the ROC curve (AUC). Most of the reward-responsive neurons were not activated following the delivery of

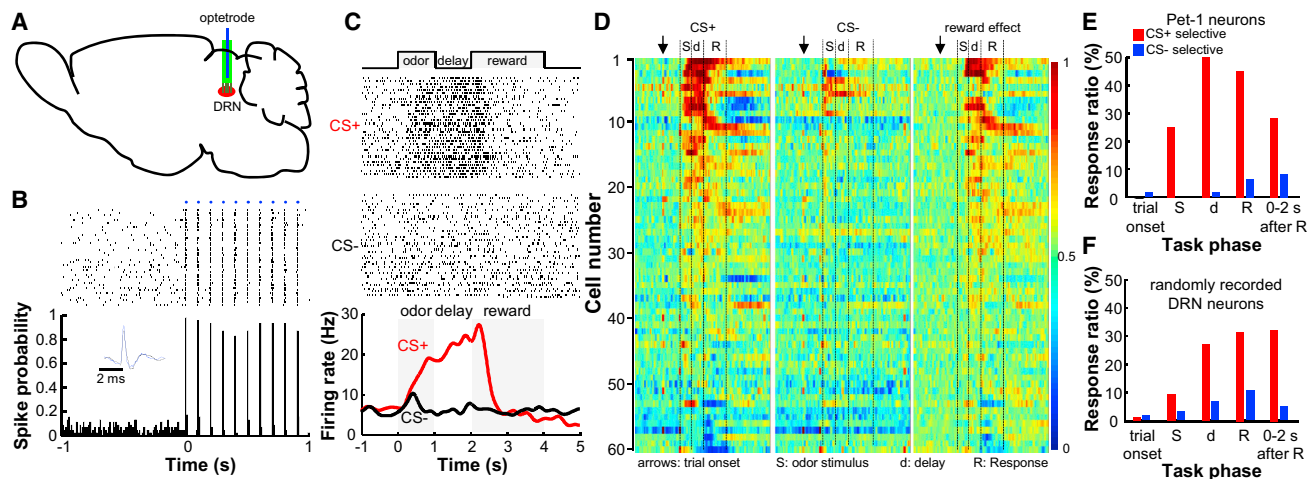


Figure 4. DRN Pet-1 Neurons Are Activated in Response to Rewarding Stimuli in an Olfactory Go/No-Go Task

(A) DRN neurons were recorded from behaving mice with optetrodes.

(B) Raster plot (upper) and peristimulus time histogram (PSTH; lower, bin width = 10 ms) show that light stimulation reliably evoked spike firing of a DRN neuron. The inset shows that light-evoked (blue) and spontaneous (black) spikes had similar waveforms.

(C) Raster plot and PSTH (smoothed with a Gaussian kernel, $\sigma = 100$ ms) of the activity of a DRN Pet-1 neuron aligned to odor onset.

(D) Population activity of DRN Pet-1 neurons in the Go/No-go task. Each row represents the activity of a single neuron. For CS+ and CS- trials (left and middle panels), firing rates were compared with the mean rates before trial onset (arrows) to calculate receiver operating characteristic (ROC) values and are represented with colors. AUC, the area under a ROC curve. An AUC value of 0.5 indicates no difference from the mean activity before trial onset. Reward effect (right panel) was computed by comparing the firing rates of CS+ and CS- trials of the same neurons and an AUC value of 0.5 indicates no selectivity.

(E) Distribution of identified DRN Pet-1 neurons with significant selective responses to CS+ or CS- within different phases of Go/No-go tasks.

(F) Distribution of response selectivity for the 159 randomly recorded DRN neurons without cell-type identification. See also Figure S4.

reward-negative cues (Figure 4D). Further analysis revealed that ~65% of the identified Pet-1 neurons (39 out of 60 cells) exhibit a significant reward effect during the period between odor cue and reward delivery.

An additional set of 159 neurons was randomly recorded from the DRN without the confirmation of optical tagging. Many of these cells showed tonic excitation only during reward-associated tasks, although the response patterns were more diverse than the identified Pet-1 neurons (Figures S4H and S4I). At the population level, the number of DRN neurons selectively recruited during reward tasks gradually increased after the onset of sensory stimuli (Figure S4J). During the specific phases of waiting and sucrose consumption, the responsive ratio reached a maximum of ~50% for Pet-1 cells and ~30% for randomly recorded DRN cells (Figures 4E and 4F). In contrast, only ~5% of DRN neurons were selectively excited when an odorant was not associated with reward (Figure 4F).

The Activity of DRN Pet-1 Neurons Changes Cortical Activation Patterns

The strong behavioral effects suggest that the activity of DRN Pet-1 neurons can exert rapid physiological influences in the motor cortex. To study whether DRN stimulation could guide the change of cortical activity at the single-neuron level, we carried out recordings from head-fixed mice undergoing an operant brain-machine interface (BMI) task. While in behavioral experiments mice received light stimulation by directly executing a specific physical movement, in the BMI task

animals learned to control laser pulses delivery into the DRN through the modulation of neuronal activity in the cortex (Koralek et al., 2012). Tetrode recordings were performed from the vibrissa motor cortical area (vM1) of ePet1-DRN^{ChR2} mice (Figures 5A and S5A). After isolating several single units from one tetrode, the ensemble activity was used to trigger light stimulation in the DRN (3 s; 20 Hz) when the firing frequency crossed a predetermined threshold during the presentation of 10 s odorant pulses (Figure 5B). To prevent neuronal runaway firing, odorant pulses were applied only after the ensemble activity was below another threshold approximately equal to the baseline level.

In this closed-loop setting, changes of firing activity from a single recording site in the cortex determined the occurrence of DRN stimulation and the subsequent reinforcement of cortical activity. Before training, the vM1 cells lacked a clear response to odor stimulation, and the mean firing frequency occasionally crossed the threshold during odorant pulses, resulting in DRN stimulation, which in turn increased the chance of threshold crossing of vM1 neurons. After training, vM1 cells exhibited significant excitation tightly coupled to the onset of odorant pulses ($p < 0.01$; permutation test; $n = 52$ recording sites from 15 mice) (Figures 5C, 5D, and S5B). Neuronal ensembles rapidly increased response strength within 20 trials, slowly reaching a peak with additional training (Figures 5E and 5F). The change in neuronal activity led to dramatic increase in the events of firing above the reward threshold only during odor pulses (Figure 5G). A majority of sorted single units (137 out of 195) exhibited significant excitatory responses after training,

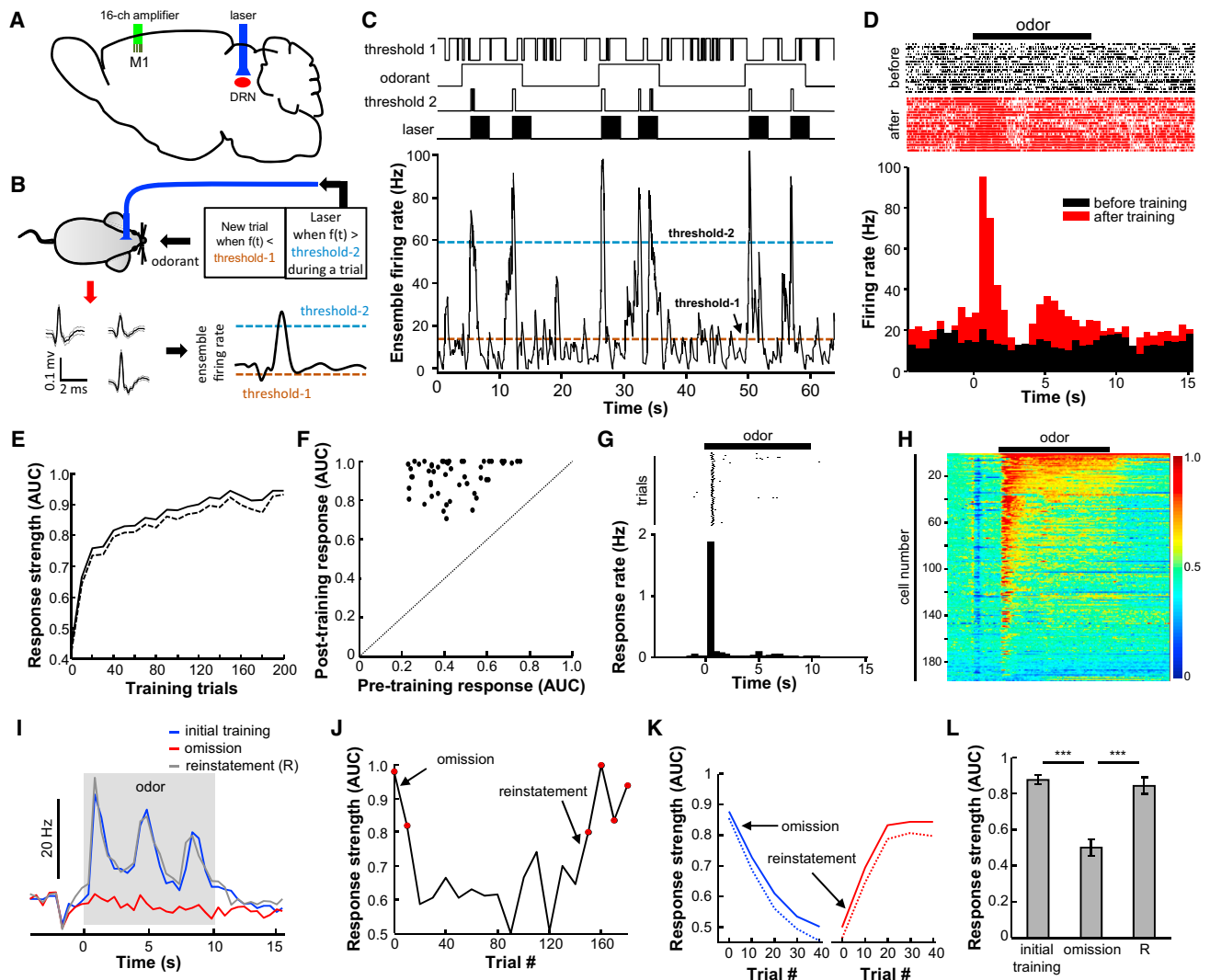


Figure 5. Phasic Activation of the DRN Pet-1 Neurons Efficiently Directs the Change of Activity Patterns of Individual Cortical Neurons

(A) Ensemble spiking activity was recorded from the vM1 of ePet1-DRN^{ChR2} mice implanted with an optical fiber over the DRN for light stimulation.

(B) Schematic for the BMI operant task. The ensemble firing rates of vM1 neurons defined odor onset and laser delivery to the DRN. Odorant pulses were applied when the ensemble firing rates were below a predetermined firing rate (threshold-1). Light pulses (3 s; 20 Hz) were generated when ensemble-firing rates were above a predefined high level (threshold-2) during odor presentation.

(C) Example traces from a well-trained vM1 ensemble. Neurons responded vigorously and reliably during odorant pulses.

(D) Raster plots and PSTH (bin = 0.5 s) showing that an ensemble lacked response to odorants before the BMI task training and responded strongly after training.

(E) Averaged learning curve of 52 well-trained ensembles. The dashed line represents mean-SEM.

(F) 2-D plot comparing ensemble response strength to the odorant before and after the BMI training.

(G) Raster plot and PSTH (bin = 0.5 s), showing the task response frequency of a well-trained vM1 ensemble. Threshold-2 crossing by the ensemble-firing rate was designated as a task response.

(H) Heatmap showing the ROC representation of PSTH data for all recorded single units (n = 195).

(I) Odor-evoked responses of one ensemble were reduced by the omission of light stimulation and recovered after stimulation reinstatement.

(J) Time-series plot of response strength showing the effect of stimulation omission and reinstatement on an ensemble. The red dots indicate significant responses ($p < 0.01$; permutation test).

(K) Population data showing the effects of stimulation omission and reinstatement across time (n = 14 ensembles from 6 mice).

(L) Group data of stimulation omission tests (** $p < 0.001$; paired t test). See also Figure S5.

demonstrating learning effects at the level of single neurons (Figures 5H and S5C).

We asked whether mice intentionally controlled M1 activity to receive the light stimulation reward (goal directed) or whether the

actions were habitually produced through previous reinforcement. As goal-directed behaviors, but not habits, are sensitively affected by the contingency of an action on reward delivery and reward expectation (Jog et al., 1999; Koralek et al., 2012), we

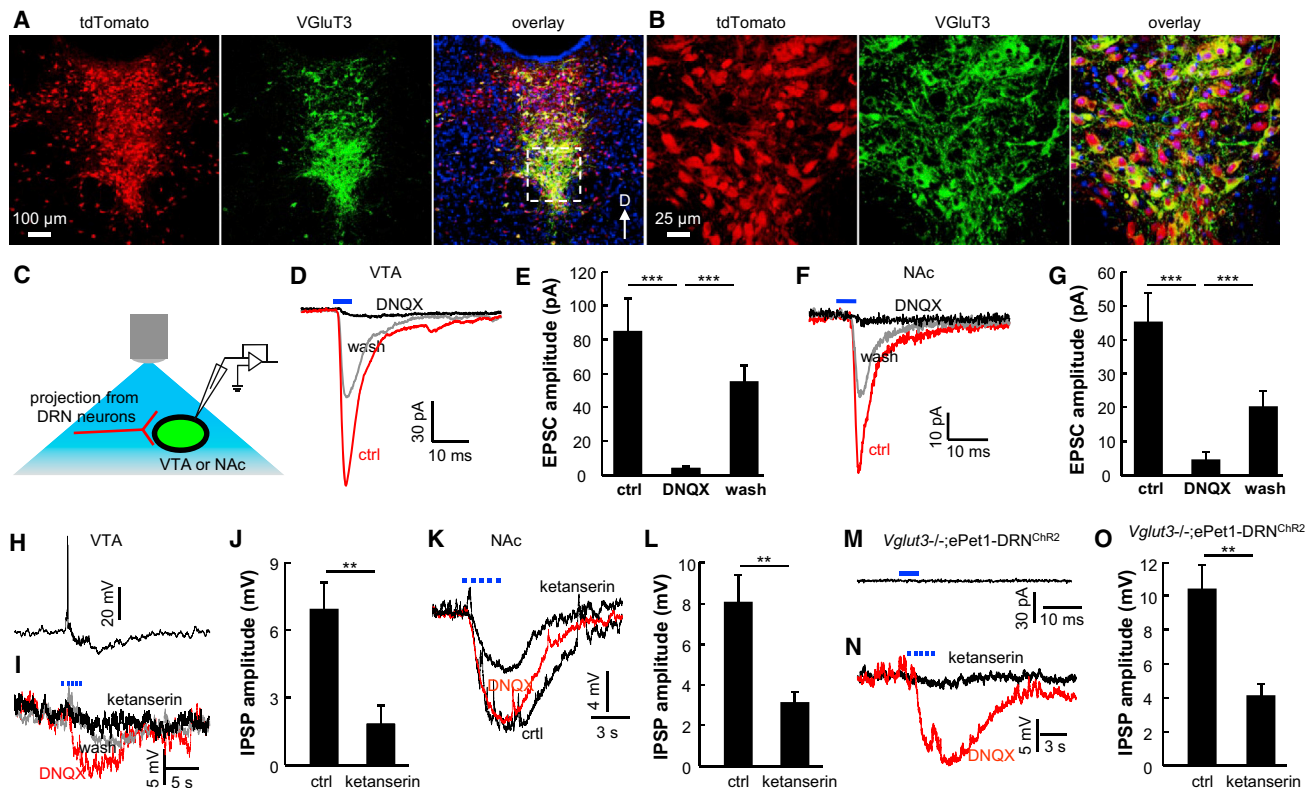


Figure 6. DRN Pet-1 Neurons Release 5-HT and Glutamate

(A and B) In an ePet1-Cre;Ai14 mouse, VGlut3 (green) is expressed in many tdTomato-labeled neurons (red) along the midline. Panels in (B) show the zoom-in view of the dashed rectangular area in (A).

(C) Schematic diagram showing the method of optogenetic stimulation and recordings from the VTA or the NAc in brain slices.

(D and E) Representative recording traces from a VTA neuron (D) and group data (E) reveal that brief light stimulation of ChR2⁺ axonal terminals produced fast EPSCs that were reversibly blocked by DNQX (**p < 0.001; paired t tests; n = 13 cells).

(F and G) Glutamatergic EPSCs were also evoked by single-pulse light stimulations in the NAc shell (**p < 0.001; paired t tests; n = 7 cells).

(H and I) Current-clamp recordings from a single VTA neuron show that trains of light pulses (3 s; 20 Hz) resulted in brief excitation, followed by slow inhibition (H). The initial excitatory response was blocked by DNQX, whereas the slow inhibitory response was largely abolished by ketanserin, which blocks 5-HT_{2A} and 5-HT_{2C} receptors (I).

(J) Group data showing the effect of ketanserin on the slow IPSPs (**p < 0.01; paired t test; n = 6 cells).

(K and L) Slow 5-HT effects were also observed in the NAc (**p < 0.01; paired t test; n = 7 cells).

(M and N) Brief light stimulation failed to elicit any fast EPSC from a cell in the VTA of a *Vglut3*^{-/-};ePet1-DRN^{ChR2} mouse (M), but repetitive light stimulation (3 s; 20 Hz) evoked slow IPSP that was largely abolished by ketanserin (N).

(O) Group data showing that the slow IPSPs were significantly reduced by ketanserin in *Vglut3*^{-/-};ePet1-DRN^{ChR2} mice (*p < 0.01; paired t test; n = 6 cells). See also Figure S6.

examined the effect of omitting stimulation after correct responses or administering light stimulation irrespective of target achievement. Both stimulation omission and contingency degradation reduced responses to the chance level after 30–40 trials, and the correct responses could be re-established after 10–20 trials for the reinstatement of stimulation coupling (Figures 5I–5L and S5D–S5G). The omission effect is odor specific. The introduction of a novel odorant lacking light stimulation did not affect responses to the previously rewarding odorant, and the new odorant was significantly less likely to evoke effective excitatory responses (Figure S5H). Thus, these results suggest that DRN activation guides goal-directed learning and can be exploited to efficiently build neuronal activation patterns in the cortex.

Stimulation of DRN Pet-1 Neurons Produces Glutamate and 5-HT Effects

Although the ePet1-Cre mouse line was commonly used to drive gene expression in central 5-HT neurons, it remained unclear whether DRN Pet-1 neurons only release 5-HT. About two thirds of 5-HT neurons in the DRN express VGlut3 (Hioki et al., 2010), a vesicular transporter that is believed to concentrate glutamate into synaptic vesicles (Fremau et al., 2004; Hioki et al., 2010). We labeled Pet-1 neurons with tdTomato in ePet1-Cre;Ai14 mice and confirmed VGlut3 expression in a substantial number of DRN Pet-1 neurons (Figures 6A and 6B). VGlut3 is essential for the release of glutamate by auditory hair cells (Seal et al., 2008), but it has not been tested whether DRN Pet-1 neurons

use glutamate as a neurotransmitter and, if so, whether the potential glutamate effect requires VGluT3.

The axonal terminals of DRN Pet-1 neurons are densely distributed in the midbrain VTA and the forebrain nucleus accumbens (NAc) shell (Figures S6A and S6B), which represent the key components in the reward system (Berridge and Kringelbach, 2008; Berridge et al., 2009). In both iClass assays and conditioned place preference (CPP) tests, we observed a strong reinforcement effect following the optical stimulation of axonal terminals in the VTA of ePet1-DRN^{Chr2} mice (Figures S6C–S6H). Terminal stimulation in the NAc was ineffective in inducing a clear learning effect in iClass assays, but this stimulation did support CPP (Figures S6F–S6I). We tested whether the reinforcement effects depended on the potential antidromic activation by inactivating DRN neurons with intracranial lidocaine injection before training sessions (Stuber et al., 2011). Terminal stimulation in the VTA and NAc remained effective to produce significant reinforcement effects in the assays of iClass or CPP following the soma inactivation in the DRN (Figures S6C–S6I), suggesting that the reward signaling of DRN Pet-1 neurons might be mediated by their axonal fibers in the VTA and NAc or axonal collaterals outside of these two target areas.

Whole-cell patch-clamp recordings were performed using brain slices of ePet1-DRN^{Chr2} mice to examine the effect of stimulating axonal terminals from DRN Pet-1 neurons on post-synaptic neurons (Figures 6C and S6J–S6M). In the VTA and NAc, single-pulse light stimulation produced fast excitatory responses that were reversibly abolished by the application of 6,7-dinitroquinoxaline-2,3-dione (DNQX), a selective glutamate receptor antagonist (Figures 6D–6G). In both areas, prolonged light stimulation (20 Hz for 3 s) typically produced slow inhibitory responses that were substantially reduced by ketanserin (Figures 6H–6L), a drug that blocks 5-HT_{2A} and 5-HT_{2C} receptors. In *Vglut3*^{−/−};ePet1-DRN^{Chr2} mice lacking VGluT3 expression (Figure S6N), we detected 5-HT effects but did not observe any fast glutamatergic EPSCs in the 43 cells tested (Figures 6M–6O). Thus, DRN Pet-1 neurons release glutamate and 5-HT as neurotransmitters, and VGluT3 is required for glutamate release.

Both Glutamate and 5-HT Contribute to Reward Signaling

Since a vast majority of DRN Pet-1 neurons are serotonergic, we studied the role of 5-HT in reward signaling by analyzing the behavioral effects after genetically or chemically depleting brain 5-HT. *Tph2*^{−/−};ePet1-DRN^{Chr2} mice lacked Tph2 expression and exhibited a dramatic reduction of 5-HT levels in the DRN (Figures S7A–S7C). In iClass training tasks with 20 Hz stimulation, knocking out the *Tph2* gene mildly reduced the center entry numbers or center duration in certain test sessions, but overall, the stimulations produced a qualitatively clear reinforcement effect (Figures 7A–7D, S7E, and S7F). Brain 5-HT concentrations were reduced to ~16% of basal levels following the injection of 4-Chloro-L-phenylalanine (L-pCPA), a tryptophan hydroxylase inhibitor (Figure S7B–S7D) (Liu et al., 2011). Depletion of 5-HT using L-pCPA similarly had only mild effects on ePet1-DRN^{Chr2} mice (Figure 7A–7D, S7E, and S7F).

We then investigated the role of glutamate through the analysis of the behavioral phenotypes of *Vglut3*^{−/−};ePet1-DRN^{Chr2} mice. In iClass assays, *Vglut3*^{−/−};ePet1-DRN^{Chr2} mice exhibited significantly fewer center entries but comparable center exploration time (Figures 7A–7D), suggesting that both 5-HT and glutamate play a role, and neither of the two transmitters is absolutely essential for the reinforcement effect of DRN Pet-1 neurons. Because both *Vglut3* and *Tph2* are located on the same chromosome, it is impossible to generate a double mutant through crossbreeding to examine the effect of disrupting both 5-HT and glutamate release. Therefore, we injected L-pCPA into *Vglut3*^{−/−};ePet1-DRN^{Chr2} mice to block the transmission of glutamate and 5-HT by DRN Pet-1 neurons. L-pCPA injection completely abolished the reinforcement effect of DRN stimulation on *Vglut3*^{−/−};ePet1-DRN^{Chr2} mice (Figures 7A–7D, S7E, and S7F).

In two-bottle preference tests without light coupling, both *Tph2*^{−/−} mice and *Vglut3*^{−/−} mice preferred sucrose in a concentration-dependent manner (Figures 7E and 7F). Coupling DRN stimulation to licking for water (20 Hz; 1 s) effectively shifted the sucrose preference of *Tph2*^{−/−};ePet1-DRN^{Chr2} mice and produced a reward value comparable to those of ePet1-DRN^{Chr2} mice, whereas knocking out the *Vglut3* gene decreased the reward value from 5% to ~1% sucrose (Figures 7G and 7H). Following L-pCPA injection, stimulation coupling became completely ineffective to shift the sucrose preference of *Vglut3*^{−/−};ePet1-DRN^{Chr2} mice (Figures 7G and 7H), suggesting that 5-HT might contribute to the residual reward value following the disruption of glutamate release.

For self-stimulation tests in which light stimulation (20 Hz; 3 s) was produced by each nose poke out of the timeout period (fixed ratio 1[FR1]), knocking out *Vglut3* alone substantially reduced the number of nose pokes, whereas *Tph2*^{−/−};ePet1-DRN^{Chr2} mice responded normally (Figures 8A and S8A). However, the *Tph2*^{−/−};ePet1-DRN^{Chr2} mice exhibited significantly weaker response rates than ePet1-DRN^{Chr2} mice when it required five or eight consecutive pokes (FR5 and FR8) to earn light stimulation (Figures 8B–8D). Most of the *Tph2*^{−/−};ePet1-DRN^{Chr2} mice abandoned nose-poking efforts after 10 min of testing (Figure S8A), suggesting that 5-HT is required for maintaining motivation during more difficult tasks.

For Go/No-go olfactory discrimination tests, DRN stimulation was much less effective to drive *Tph2*^{−/−};ePet1-DRN^{Chr2} mice and *Vglut3*^{−/−};ePet1-DRN^{Chr2} mice to perform the task. Both groups of mutant mice required significantly longer time to initiate a new trial (Figure 8E). In contrast to ePet1-DRN^{Chr2} mice that completed at least 500 trials within a daily 240 min training session, *Tph2*^{−/−};ePet1-DRN^{Chr2} mice performed a maximum of 140 trials and abandoned their efforts after 150 min (Figure 8F). *Vglut3*^{−/−};ePet1-DRN^{Chr2} mice maintained a lower rate of response throughout the session and completed ~300 trials. Despite more variable and slower learning, both *Tph2* and *Vglut3* mutant mice achieved a correct ratio of ~85% and ~75%, respectively (Figure 8G, S8B, and S8C). L-pCPA injection into *Vglut3*^{−/−};ePet1-DRN^{Chr2} mice completely blocked the learning effect induced by DRN stimulation (Figure 8G).

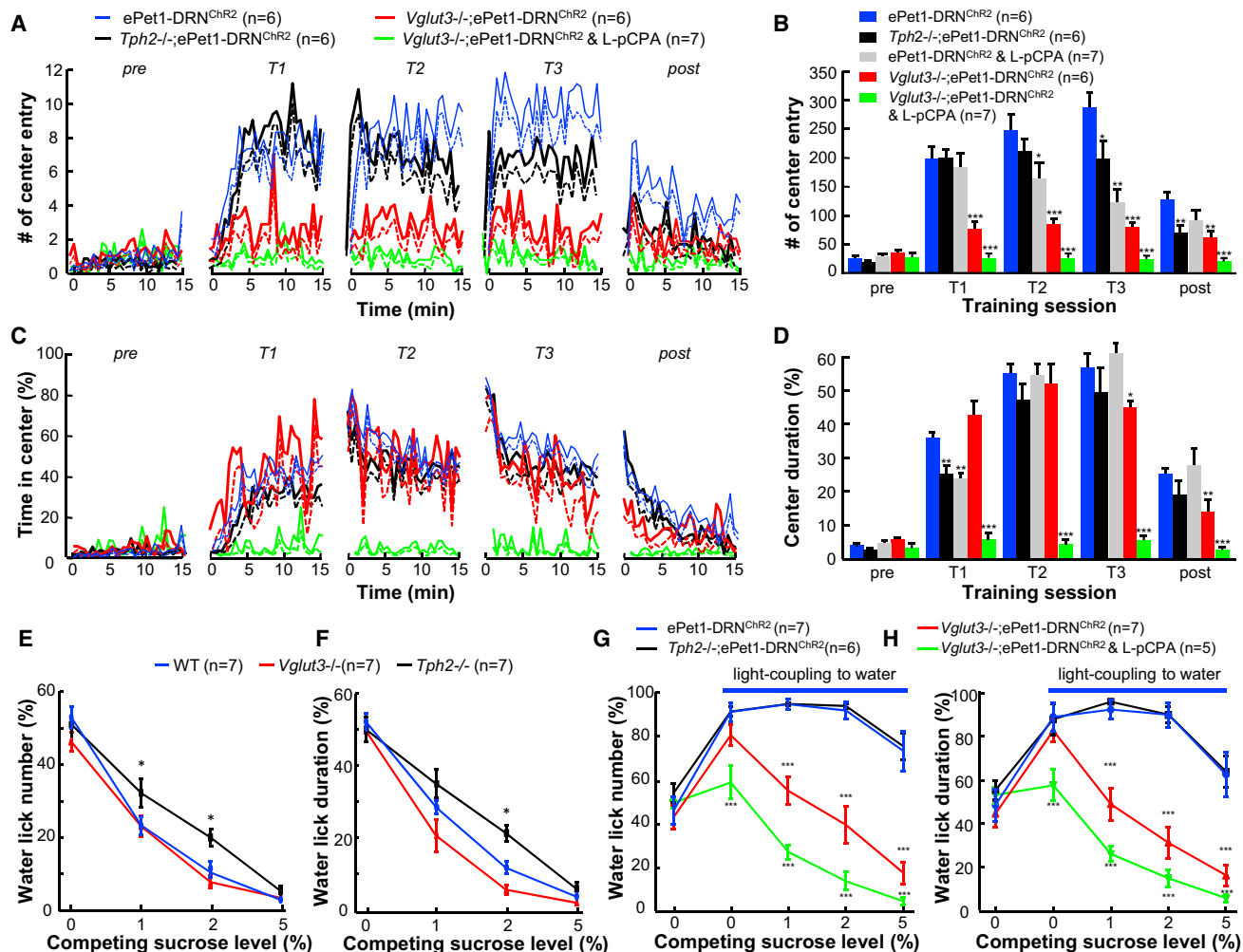


Figure 7. Data from iClass Tests and Two-Bottle Preference Tests Reveal that Both 5-HT and Glutamate Contribute to Reward Signaling by DRN Pet-1 Neurons

(A and B) In iClass tests, *Tph2*^{-/-};ePet1-DRN^{ChR2} mice and L-pCPA-treated ePet1-DRN^{ChR2} mice showed a mild but statistically significant reduction in the center entry number for certain training sessions (T2 or T3). *Vglut3*^{-/-};ePet1-DRN^{ChR2} exhibited ~50% reduction in the number of center entries of all training sessions. L-pCPA injection into *Vglut3*^{-/-};ePet1-DRN^{ChR2} mice completely abolished the reward effect produced by the activation of DRN Pet-1 neurons. **p* < 0.01; ****p* < 0.001; *t* tests between test groups and ePet1-DRN^{ChR2} control mice.

(C and D) The effect of knocking out the *Vglut3* gene and/or depleting 5-HT on the center duration.

(E and F) The sucrose preference scores quantified with lick numbers and lick duration, respectively. Both *Tph2*^{-/-} and *Vglut3*^{-/-} mice preferred sucrose to water, but the sucrose preference scores of *Tph2*^{-/-} mice were lower than those of WT mice at the concentrations of 1% and 2%. **p* < 0.05; two-way ANOVA and then Dunnett's multiple comparison tests between mutants and WT.

(G and H) Sucrose preference scores show that light stimulation of the DRN Pet-1 neurons in *Vglut3*^{-/-};ePet1-DRN^{ChR2} mice produced a reward value of ~1% sucrose. L-pCPA injection into these mice completely disrupted reward signaling. ***p* < 0.01; ****p* < 0.001; one-way ANOVA and then Tukey's post hoc test between test groups and ePet1-DRN^{ChR2} control mice. See also Figure S7.

DISCUSSION

In this study, we investigated whether DRN neurons signal reward or punishment. Multiple behavioral assays reveal that the optogenetic activation of DRN Pet-1 neurons strongly reinforces behavior and efficiently guides learning. Recordings from behaving mice show that DRN Pet-1 neurons increase activity during reward-associated tasks. In addition, the stimulation of these neurons rapidly changes the activation pattern of cortical neurons to establish the predictive association of

neuronal activity with a specific sensory stimulus. In slice preparations, the stimulation of DRN Pet-1 neurons produces postsynaptic effects mediated by 5-HT and glutamate receptors. In addition, the reward signaling is disrupted in mice lacking functional *Tph2* and *Vglut3*. These experiments demonstrate that these DRN Pet-1 neurons encode reward and 5-HT and glutamate contribute to these effects.

Since the pioneering intracranial electric self-stimulation experiments of Olds and Milner (1954), a set of brain structures have been identified as important for reward processing. The

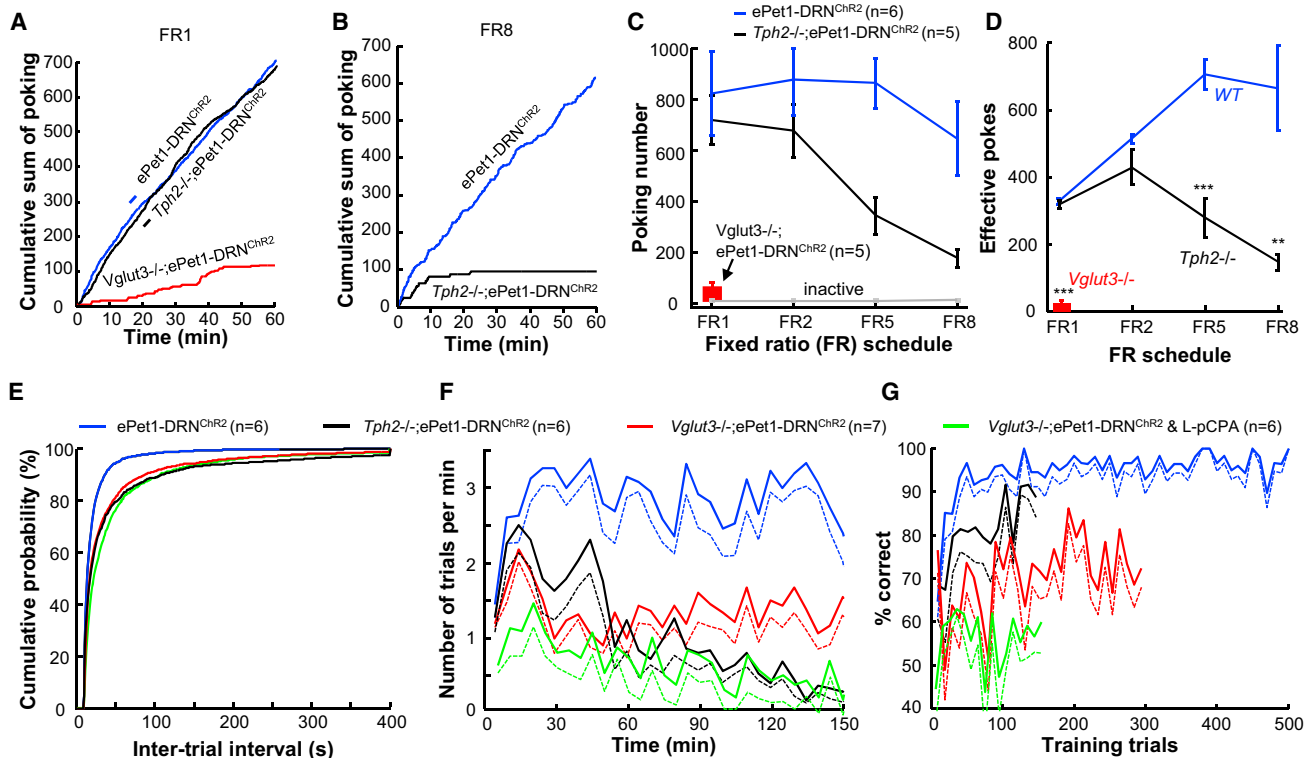


Figure 8. Mice Lacking *Tph2* or *Vglut3* Show Impaired Acquisition of Self-Stimulation and Olfactory Discrimination Learning

(A–D) The behavioral phenotypes of *Tph2* and *Vglut3* mutant mice in the tests of light self-administration. *Vglut3*^{-/-};ePet1-DRN^{ChR2} mice, but not *Tph2*^{-/-};ePet1-DRN^{ChR2} mice, exhibited a dramatic decrease in nose poking in tests involving an FR1 schedule (A). *Tph2*^{-/-};ePet1-DRN^{ChR2} mice responded with much lower intensity than *Tph2*^{+/+};ePet1-DRN^{ChR2} mice in tests involving FR5 and FR8 schedules (B–D). In (D), a poke is considered effective if it occurred outside of the timeout period. ***p* < 0.01; ****p* < 0.001; *t* tests between test groups (*Tph2*^{-/-} or *Vglut3*^{-/-}) and the ePet1-DRN^{ChR2} control group.

(E–G) Knocking out either the *Tph2* gene or the *Vglut3* gene disrupted the olfactory discrimination learning directed by the stimulation of DRN Pet-1 neurons.

(E) Plots of cumulative probability against intertrial intervals for different animal groups. Knocking out the *Tph2* or *Vglut3* gene significantly increased the time required to initiate a new trial during the Go/No-go olfactory discrimination test (*p* < 0.001; Kolmogorov-Smirnov test between knockout and WT mice).

(F) Plots of trials per minute for different animal groups engaged in olfactory Go/No-go tasks driven by DRN stimulation.

(G) The learning curves of different test groups. The plot for ePet1-DRN^{ChR2} mice is derived from Figure 3C. See also Figure S8.

reward “hotspots” include the midbrain VTA, the NAc shell, the ventral pallidum, the hypothalamus, the anterior cingulate cortex, and the orbitofrontal cortex (Berridge and Kringelbach, 2008). Neuronal activities in these areas might be integrated to process distinct components of reward, such as hedonia (“liking”), motivation (“wanting”), and learning (Berridge et al., 2009). Our behavioral assays show that the optogenetic activation of DRN Pet-1 neurons produces incentive motivation that promotes vigorous self-stimulation and generates high reward value comparable to that of ingested sucrose. In addition, the activation of DRN Pet-1 neurons efficiently guides sensory discrimination learning. These behavioral assays examine both operant and Pavlovian conditioning. The strong behavioral effects in all tests suggest that the DRN is a reward center and should be incorporated into models of the brain reward system.

DRN Pet-1 neurons might signal reward by targeting multiple brain areas, including the established reward centers. In addition to projections to sensory and motor cortical areas, the DRN forms extensive interconnections with essentially all nodes in the reward system (Vertes, 1991; Watabe-Uchida et al., 2012).

The effectiveness of stimulating axonal terminals in the VTA and NAc suggests that DRN projections to these two brain areas are involved in reward signaling. However, the particularly strong effects of VTA stimulation do not necessarily indicate that DRN Pet-1 neurons signal reward solely through midbrain dopamine neurons. First, stimulating axonal terminals in the NAc could also reinforce animal behaviors in CPP tests. Moreover, DRN Pet-1 neurons and VTA dopamine neurons have different activation patterns in reward-associated tasks. After learning, dopamine neurons fire transiently following a reward-predicting cue (Cohen et al., 2012; Schultz et al., 1997), whereas DRN Pet-1 neurons exhibit a tonic increase in firing activity until reward delivery. This firing pattern has also been reported for subsets of randomly recorded DRN neurons (Bromberg-Martin et al., 2010; Miyazaki et al., 2011; Nakamura et al., 2008). The distinct activation patterns suggest that DRN Pet-1 neurons and VTA dopamine neurons might play different roles in reward processing. Although dopamine has been synonymous with reward, previous studies have also revealed dopamine-independent reward behaviors. For example, sucrose remains rewarding for mice

without dopamine (Cannon and Palmiter, 2003), and cocaine can produce reward through the 5-HT signaling pathway (Sora et al., 2001). Anatomically, there are strong reciprocal projections between the DRN and VTA (Kalén et al., 1988; Vertes, 1991; Watabe-Uchida et al., 2012). Thus, DRN Pet-1 neurons may target VTA dopamine neurons as well as other brain areas to organize reward behaviors.

The fast learning rates observed in the operant behavioral tests and the BMI-based tasks suggest that DRN neurons can rapidly modify neural circuits and facilitate animal learning. Coupling the stimulation of DRN Pet-1 neurons to the operant action of neuronal activity in the motor cortex, we observed real-time changes in the neuronal activity patterns of mice engaged in BMI tasks. This approach might be used to facilitate learning and memory formation. To control the movement of an external object with brain activity, neuroprosthetics accomplish the challenging task of decoding movement intention through long-term cortical recordings using a large number of electrodes (Koralek et al., 2012; Taylor et al., 2002). Our BMI recordings suggest that the method of coupling DRN stimulation to a given pattern of neuronal activation can increase the efficiency of establishing the neural code for controlling neuroprosthetic devices.

Another key finding of this study is that both 5-HT and glutamate contribute to the reward signaling of DRN Pet-1 neurons. Although DRN Pet-1 neurons have been considered serotonergic, these cells release glutamate and 5-HT. Blocking 5-HT synthesis or glutamate release results in partial impairments in a task-dependent manner. For behavioral tests of iClass, two-bottle preference, and self-stimulation with the FR1 schedule, knocking out *Vglut3* but not *Tph2* produces more obvious impairments. For *Tph2* knockout mice, a clear performance reduction is observed when more efforts and longer intervals are required for earning DRN stimulation, such as in self-stimulation assays involving FR5 and FR8 schedules and olfactory Go/No-go tests. 5-HT might be particularly important for maintaining motivation in response to difficult tasks. Most importantly, the reward effects of stimulating DRN Pet-1 neurons are completely eliminated through the injection of L-pCPA, a tryptophan hydroxylase inhibitor, into VGLUT3 knockout mice, suggesting an intersection of the two neurotransmission pathways. Thus, our data from mutant mice and L-pCPA injections strongly suggest that both 5-HT and glutamate contribute to the reward signaling of DRN Pet-1 neurons. Deficits in DRN 5-HT neurons have been implicated in depression of humans and animals (Amat et al., 2005; Krishnan and Nestler, 2008; Mann, 1999; Warden et al., 2012), a core symptom of which is anhedonia (Der-Avakian and Markou, 2012). Our data suggest that both 5-HT and glutamate signaling pathways of DRN neurons could be intervened to manipulate reward processing and treat anhedonia.

Several caveats of our approaches need to be mentioned. We examined the role of 5-HT by genetically and chemically depleting 5-HT. The contribution of glutamate was studied by analyzing the behavioral phenotypes of knocking out the *Vglut3* gene. We find that VGLUT3 is required for the glutamatergic effects of DRN Pet-1 neurons and its mutant exhibits impairments in reward behaviors elicited by selective stimulation of these neurons. However, data from knockout mice might be

confounded by developmental compensation. In addition to neurons in the raphe, hair cells in the ear and subsets of neurons in the striatum and cortex express VGLUT3 (Herzog et al., 2004; Seal et al., 2008). *Vglut3*^{-/-} mice exhibit normal locomotor behavior but have deafness and seizure-like electrical activity in the cortex (Seal et al., 2008), suggesting limitations of using these mice to precisely study the functions of glutamate from DRN Pet-1 neurons in reward processing. Clean dissection of the roles of 5-HT and glutamate might be achieved by temporally and spatially controlled conditional knockout of either the *Tph2* or *Vglut3* gene in the DRN.

How can our findings be reconciled with the published hypotheses that the activity of DRN 5-HT neurons encodes punishment? Both the DRN and the medial raphe nucleus (MRN) contain 5-HT neurons, and the DRN is further separated into different subdivisions based on neurotransmitter phenotypes. For example, VGLUT3 is expressed in 5-HT neurons in the center, but not in the two lateral wings of the DRN (Hioki et al., 2010). Since we mainly stimulated neurons in the center of the DRN, it cannot be excluded that some 5-HT neurons in the lateral wings of the DRN and the MRN may encode punishment signals (Lechin et al., 2006). Nevertheless, the punishment theory is largely based on the experiments of prolonged manipulations of global 5-HT levels, which differ from the phasic activation of DRN neurons in terms of temporal and spatial scales. In addition, it had not been firmly established that DRN neurons can release glutamate, and early studies neglected the contribution of glutamate. Although the DRN has been reported to be an effective locus that supports electric self-stimulation, the ineffectiveness of 5-HT depletion has led to the suggestion that the reinforcement effect is produced through the stimulation of fibers of passage rather than neurons in the DRN (Deakin, 1980; Simon et al., 1976). While the reasons for these discrepancies between the behavioral studies are unclear, physiological studies have also challenged the simplified view that DRN neurons inhibit dopamine neurons. The optogenetic stimulation of DRN terminals directly excites VTA neurons through the action on glutamate receptors (Figure 6). Recordings in vivo show that 5-HT can exert complex excitation and inhibition patterns in dopamine neurons (Gervais and Rouillard, 2000). The effects of 5-HT and glutamate can be mediated by many receptors at both presynaptic and postsynaptic sites (Conn and Pin, 1997; Dingledine et al., 1999; Hoyer et al., 2002), suggesting a rich repertoire of physiological functions through DRN Pet-1 neurons. Analyzing the roles of these neurons in various microcircuits within the reward system might provide further insights into cellular and circuit mechanisms of reward processing.

EXPERIMENTAL PROCEDURES

Methods and materials are described in details in [Supplemental Experimental Procedures](#).

All experiments were performed on adult mice (10–16 weeks old, both male and female). The three transgenic mouse lines (ePet1-Cre, *Tph2*^{-/-}, and *Vglut3*^{-/-}) were crossed to the genetic background of C57BL/6N (Vitalriver Laboratory Animals, Beijing). The *Tph2*^{-/-};ePet1-Cre and *Vglut3*^{-/-};ePet1-Cre lines were produced by crossing ePet1-Cre mice with *Tph2*^{-/-} and *Vglut3*^{-/-} mice, respectively. For transgene expression, adeno-associated viral particles of serotype 9 for AAV-DIO-ChR2-mCherry or AAV-DIO-mCherry

were infused into the DRN of ePet1-Cre mice. The mice were allowed to recover for 2–3 weeks before behavioral assays or physiological recordings. All procedures were conducted with the approval of the institutional Animal Care and Use Committee of NIBS, Beijing in accordance with governmental regulations of China.

We carried out five different behavioral tests to investigate the reinforcement effect of activating DRN Pet-1 neurons. Blue light pulses, generated by a diode-pumped solid-state laser, were passed to the DRN through an implanted optical fiber (473 nm wavelength, 15 ms pulse duration, and 5 or 20 Hz frequency; 20 mW output power measured at the fiber tip with continuous light output; fiber diameter = 200 μ m and NA = 0.22). The so-called iClass task was used to examine whether mice could be reinforced to explore a pre-designated area in an open field by coupling optical stimulation of DRN neurons with the mouse behavior of exploring this specific area. Animal positions were monitored by an overhead camera and laser was controlled by a custom-written Matlab program. Two-bottle preference tests were performed to examine whether mouse sucrose preference could be shifted by coupling DRN stimulation with animal licking for water (473 nm, pulse duration 15 ms, and 20 Hz for 1 s or 5 Hz for 2 s). The preference scores were calculated as the ratio of the number or duration of licks of the light-coupled water bottle to the total lick number or duration within test sessions. The tests of intracranial optical self-stimulation were carried out by placing mice in an operant chamber equipped with two nose-poke detectors (one "active hole" and one "inactive hole"). Nose-poking through the active hole resulted in the delivery of blue light pulses into the DRN through the optical fiber. We examined the effects of photoactivating DRN Pet-1 neurons on learning by subjecting the animals to an olfactory Go/No-go learning paradigm. Mice were trained to touch a lickometer for the reward of sucrose solution or DRN stimulation following the presentation of a specific odorant. Mice needed to inhibit licking following the presentation of another odorant to avoid the punishment of timeout. We used the standard three-chamber unbiased CPP test to evaluate the reward effect of stimulating ChR2⁺ terminals in the VTA or the NAc. To chemically deplete 5-HT, mice were intraperitoneally injected with L-pCPA twice a day for 3 consecutive days before being subjected to behavioral tests. Brain monoamines were measured using high-performance liquid chromatography coupled with electrochemical detection.

The methods of slice preparation, whole-cell patch recording, and photostimulation are similar to those described elsewhere (Ren et al., 2011). Briefly, coronal or horizontal brain sections (300 μ m thick) were acutely prepared and continuously superfused with artificial cerebrospinal fluid (aCSF). Neurons in the DRN, VTA, or NAc shell were identified with mCherry fluorescence and DIC microscopy. Voltage-clamp and current-clamp recordings were carried out using a computer-controlled amplifier (MultiClamp700B; Molecular Devices). Blue light pulses were delivered by an optical fiber with its tip submerged in aCSF. Drugs (10 μ M DNQX, 50 μ M picrotoxin, and 10 μ M ketanerin; all from Sigma) were added to the superfusion aCSF through the dilution of stock solutions.

To record DRN neurons from mice performing olfactory discrimination tasks, water-deprived mice were head fixed above a spherical treadmill supported by floating air. After the completion of training with the Go/No-go paradigm, we recorded extracellular spiking signals with 16-channel optrodes comprising four tetrodes and an optical fiber (100 μ m diameter). The significance of neuronal response strength and selectivity was determined with permutation tests using 1,000 bootstrap replicates, and $p < 0.01$ was considered statistically significant (Ranade and Mainen, 2009). To record from the M1 cortical area from mice undergoing a BMI task, we inserted the 16-channel tetrodes into the vM1 area of head-fixed mice and placed an optical fiber above the DRN. Blue light pulses were delivered into the DRN only when the ensemble firing rates from one tetrode crossed a preset threshold during the presence of an odor. The response strengths were calculated by comparing neuronal activity during odorant application to the baseline, and their statistical significance were quantified with permutation tests.

For histology and immunohistochemistry, mice were deeply anesthetized with an overdose of pentobarbital and perfused intracardially with 0.1 M phosphate buffer saline, followed by 4% paraformaldehyde. After cryoprotection, coronal sections (35 μ m thickness) were incubated with primary rabbit antibodies against 5-HT, Thp2, VGluT3, or TH and then Cy2-conjugated

secondary antibodies. Fluorescent signals were collected using a confocal microscope (LSM510 Meta, Zeiss).

SUPPLEMENTAL INFORMATION

Supplemental Information includes eight figures, three movies, and Supplemental Experimental Procedures and can be found with this article online at <http://dx.doi.org/10.1016/j.neuron.2014.02.010>.

AUTHOR CONTRIBUTIONS

Z.L., J.Z., Y.Li., F.Hu., Y. Lu, M.M., Q.F., J.Zhang, D.W., J. Zeng, J.B., and M.L. conceived the experiments. Z.L., J.Z., Y.Li., F. Hu, and M.L. analyzed the results. M.L. wrote the paper. J.K., Z.C., and S.E.M. contributed reagents.

ACKNOWLEDGMENTS

We thank Evan S. Deneris (Case Western Reserve University) for ePet1-Cre mice, K. Deisseroth (Stanford University) for AAV-DIO-ChR2-mCherry construct, D. Duan (University of Missouri) for advice on AAV virus preparation, and J. Li (Beijing Institute of Pharmacology and Toxicology) for the analysis of brain monoamine levels. M.L. is supported by grants from the China Ministry of Science and Technology 973 Program (2010CB833902 & 2012CB837700). Z.-F.C. is supported by a NIAMS grant (AR056318).

Accepted: December 5, 2013

Published: March 19, 2014

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